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The detoxifying effects of structural elements of persimmon tannin on Chinese cobra phospholipase A₂ correlated with their structural disturbing effects well

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ABSTRACT

The effects of persimmon tannin (PT) characteristic structural elements on *Naja atra* phospholipase A₂ (PLA₂)-induced lethality, myotoxicity, and hemolysis in mice models were determined. In addition, methods including surface plasmon resonance, dynamic light scattering, and Fourier transform infrared spectroscopy were explored to uncover the possible detoxifying mechanisms of PT on snake venom PLA₂. Our results revealed that PT characteristic elements (EGCG, ECG, A-type EGCG dimer, and A-type ECG dimer) could neutralize the lethality, myotoxicity, and hemolysis of PLA₂. Moreover, the detoxifying effects of the four structural elements correlated with their structural disturbing effects well. Our results proved that A-type EGCG dimer and A-type ECG dimer may be structural requirements for the detoxifying effects of PT. We propose that the high affinity of A-type EGCG dimer and A-type ECG dimer for PLA₂ and the considerable spatial structural disturbance of PLA₂ induced by the dimers may be responsible for their antilethality, antimyotoxicity, and antihemolysis on Chinese cobra PLA₂ *in vivo*.

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1. Introduction

Phospholipase A₂ (PLA₂, E.C. 3.1.1.4) from snake venoms displays numerous toxicities, such as neurotoxicity, cardiotoxicity, myotoxicity, edema, hemolysis, and anticoagulation [1]. Besides the well-known used antisera and chemical antidotes,

plant polyphenols such as rosmarinic acid, myricetin, and aristolochic acid have been reported to be effective inhibitors of snake venoms [2]. Persimmon tannin (PT), which has a unique structure of high polymerization, high galloylation, and with both A and B type linkages and an unusual flavonol terminal unit [3], was reported to exhibit significant effects on improving the survival rate of mice injected with snake venom [4].

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Although the significant inhibition of PT on snake PLA₂ was confirmed in our previous study [5], and PT was also found to have higher affinity for PLA₂ than for BSA at physiological pH [6], detailed detoxifying mechanisms of PT were poorly understood because of the very complex structure of PT. To illustrate the antsnake PLA₂ mechanism of PT, the characteristic structural elements of PT were studied. The results showed that PT characteristic structural elements such as epigallocatechin-3-gallate-(4 β →8, 2 β →O→7)-epigallocatechin-3-gallate (A-type EGCG dimer) and epicatechin-3-gallate-(4 β →8, 2 β →O→7)-epicatechin-3-gallate (A-type ECG dimer) could potentially inhibit the catalytic activity of PLA₂ *in vitro* [7]. However, it was reported that there was a dissociation of pharmacological effects and catalytic activity in snake PLA₂s [1], so changes in the enzymatic activity of PLA₂ *in vitro* could not represent the changes in toxicity of PLA₂ *in vivo*. In addition, considering that the detoxifying process of polyphenol on PLA₂ *in vivo* is a complex process, the structural disturbing effects of PT on PLA₂ obtained from a very short interaction period could not imitate the situation *in vivo*, which may need several to dozens of hours. Considering that ECG and EGCG are the main structural elements of PT, and A-type EGCG dimer and ECG dimer are characteristic structural elements of PT, in the present study, we therefore further compared the detoxifying effects of the four structural elements (Figure 1) on PLA₂ by determining the lethality, myotoxicity, and hemolysis of PLA₂ in mice models. Moreover, more methods including surface plasmon resonance (SPR), dynamic light scattering (DLS), and Fourier transform infrared spectroscopy (FT-IR) were used to explore their interaction after incubating the mixture for 24 hours. We believe that our current work extends our previous work and provides more detailed information on the action between PLA₂ and PT structural elements, which helps to improve our understanding of the detoxifying mechanism of PT *in vivo*.

2. Methods

2.1. Materials

Chinese cobra (*N. naja atra*) phospholipase A₂ (PLA₂, 9001-84-7) was purchased from ZhongXin DongTai Nano Gene Biotechnology Co. Ltd. (Laiyang, China) with a purity of more than 95% and a molecular weight of 13,260 Da. EGCG (purity ≥95%) and ECG (purity ≥95%) were purchased from Yuanye Bio-Technology Co. Ltd. (Shanghai, China). EGCG dimer (purity ≥90%) and ECG dimer (purity ≥90%) were separated from persimmon as we previously reported [3] and further purified by preparative high performance liquid chromatography (HPLC). Their purity and identity were confirmed by HPLC and mass spectrometry. All other chemicals were of analytical reagent grade, and Milli-Q water was used in all experiments.

2.2. Animals

Male Kunming mice (19 ± 2.13 g) were purchased from the Experimental Animal Center of Disease Prevention and Control of Hubei province (Wuhan, China). All animals were maintained under standard conditions (temperature, 22–25°C; humidity, 50–70%). All experiments were performed

in compliance with the Functional evaluation program of health food and Chinese legislation on the use and care of laboratory animals.

2.3. Antilethality of PT structural elements

The median lethal dose (LD₅₀) of PLA₂ was determined by injecting different concentrations (25, 15, 10, 5, 2.5 mg/kg body weight) of venom at a constant volume of 0.2 mL/10 g body weight by intraperitoneal (i.p.) administration, and LD₅₀ was calculated according to the method described by Theakston and Reid [8]. PLA₂ was incubated with or without PT characteristic elements (the molar ratio of PLA₂/phenol was 1:15) in pH 7.4 Tris–HCl buffer at 37°C for 24 hours. The ability of PT elements to inhibit the lethal action of PLA₂ was assessed by i.p. administration of 2LD₅₀ of PLA₂ into groups (*n* = 10) of mice after starvation treatment for 12 hours. Meanwhile, the corresponding dose of PT elements was independently administered in the same manner. In all cases, the mice were observed for symptoms/signs of neurotoxicity, and the number of dead mice in each group was recorded.

2.4. Antimycrosis activities of PT structural elements

The potential ability of PT characteristic elements to inhibit PLA₂-induced myonecrosis was studied by intramuscular administration of 5 mg/kg body weight PLA₂ (incubated with or without PT elements at a molar ratio of 1:15 for 24 hours) to groups of mice (*n* = 6). The saline and corresponding dose of PT elements were independently administered in the same way. After 2 hours of administration, whole blood was collected from the mice eyes and allowed to clot for 1 hour. Later, the blood was centrifuged, and the serum was collected for experiment. CK kit (Nanjing Jiancheng) was used to determine creatine kinase (CK) levels in the mice serum.

2.5. Antihemolysis activities of PT structural elements

Indirect hemolysis was determined using the method described by Sharp et al [9] with several modifications. Briefly, the emulsified lecithin and 1 mg/mL PLA₂ (incubated with or without PT elements at a molar ratio of 1:15 for 24 hours) were incubated for 15 minutes at 37°C, then EDTA was added to terminate the interaction. The corresponding dose of PT characteristic elements, Tris–HCl buffer (negative control), and 1% Triton X-100 (positive control) were conducted in the same way. Then, 5% (v/v) mice erythrocytes were mixed with the above reaction system and incubated for 60 minutes at 37°C, followed by being centrifuged at 1500g for 10 minutes. The amount of hemoglobin released in the supernatant was estimated at 545 nm.

2.6. Surface plasmon resonance

PlexArray HT Biacore (Plexera, US) was used for SPR analysis. PT characteristic elements were printed in a photo-crosslinked chip by a photo-crosslinker system. The chip was washed with DMF, C₂H₅OH, and Milli-Q water and dried with nitrogen flow. Different concentrations (125nM, 250nM, 500nM, 1000nM, and 2000nM) of PLA₂ were used as analyte in

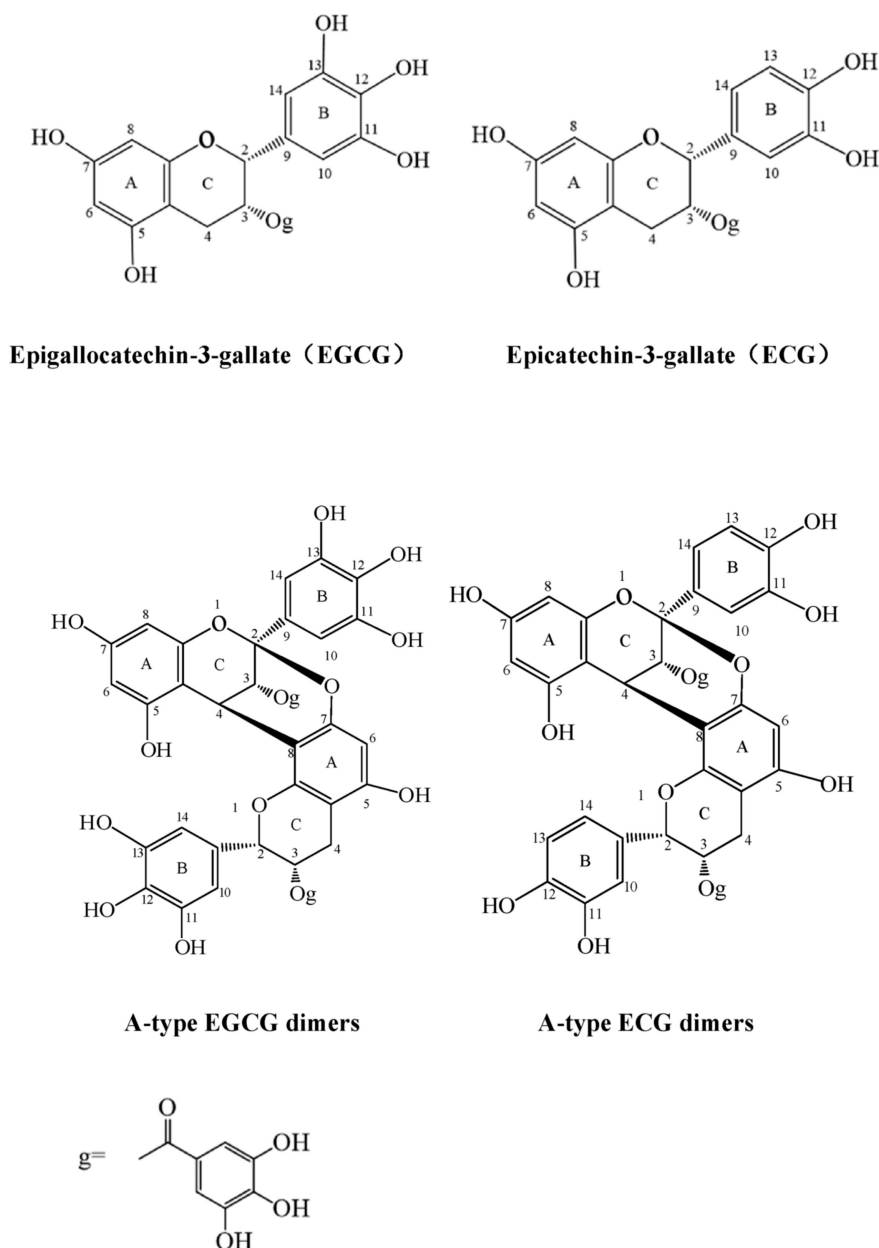


Figure 1 – Structures of the characteristic elements of persimmon tannin.

the fluid phase, with 50mM Tris–HCl (pH 7.4) as the running buffer. Both association and disassociation were performed at a flow rate of 2 $\mu\text{L/s}$ for 300 seconds under 37°C. Then, 0.5% sodium dodecyl sulfate was used to regenerate the surface and remove bound proteins from the small molecules, enabling the sensor chip to be reused for additional analyte injections.

2.7. Dynamic light scattering

The particle size and zeta potential of PLA₂ were determined by DLS using Zetasizer Nano-ZS90 (Malvern Instruments Ltd., Malvern, UK). First, 0.7 mg/mL PLA₂ was incubated with PT elements in different molar ratios (PLA₂/phenol = 1:1, 1:10,

1:20, 1:30, 1:40, 1:50) at 37°C for 24 hours. Then size measurements were undertaken at 37°C in a quartz cell. The scattering angle was 173° backscatter. The refractive index of protein (1.45) and water (1.33) at 37°C was taken from the Zetasizer software.

2.8. Fourier transform infrared spectroscopy

The infrared spectra of PLA₂ was measured using a Nicolet Nexus 470 Fourier transform spectrophotometer (Nicolet, US) in the 400–4000 cm^{-1} range, with a resolution of 4 cm^{-1} . KBr was used as a reference. For each measurement, 128 scans were taken. PLA₂ (2.7 mg/mL) was incubated with PT elements at a molar ratio of 1:15 at 37°C for 24 hours, then the mixture

was dialyzed and lyophilized prior to the experiment. The corresponding dose of PT elements was conducted in the same way. The spectra of PLA₂-PT structural units were obtained by subtracting the spectrum of PT structural units-free form from that of PLA₂-PT structural unit complexes. The curve-fitting of 1700 to 1600 cm⁻¹ (amide I) was performed using PeakFit 4.12 Software. The assignment of the bands in amide I region was in reference to Byler and Susi [10]. The peak area of each band was computed to calculate the contents of the secondary structures in PLA₂.

2.9. Statistical analysis

The results were expressed as the mean \pm standard deviation, and statistical significance between the groups was determined using one-way analysis of variance test by IBM SPSS Statistics 19.0. A *p* value less than 0.05 was considered statistically significant.

3. Results

3.1. Antilethality activities of the four structural elements

The LD₅₀ (i.p. injection) of PLA₂ calculated from the dose–response curve was 9.690 mg/kg body weight. As shown in Table 1, challenging the mice with 2LD₅₀ dose of PLA₂ (i.p. injection) caused 100% lethality, and all mice died within 2–6 hours after injection. However, PT characteristic elements increased both the survival time and survival number of mice (Table 1). EGCG dimer and ECG dimer exhibited more potent antilethality than monomers, with 60% and 50% of survival rate, respectively, whereas only 20% and 10% of survival rate for EGCG and ECG, separately, were observed. It was observed that the mice that received PLA₂ alone showed typical toxic signs of tachypnea, hemiplegia, tremble, and akinesia, whereas all the surviving mice administered with PLA₂ that was incubated with PT characteristic elements developed

none of these signs, and PT characteristic elements alone showed no lethal and neurotoxic toxicity in mice.

3.2. Antimyonecrosis activities of the four structural elements

The concentration of serum CK increased when there was necrosis in muscles [2]. Therefore, CK can be used as an enzymatic index of cellular myonecrosis damage. As shown in Figure 2, PT structural units did not induce myonecrosis as evidenced by the similar levels in CK enzyme activity with the normal group, whereas PLA₂ alone increased the CK level to about five times that of the normal group, indicating the high myotoxicity of PLA₂. However, PT characteristic elements significantly reduced the increased serum CK levels of PLA₂ poisoned mice, highlighting that PT structural units could significantly inhibit the myotoxicity of PLA₂. EGCG dimer and ECG dimer decreased the serum CK level of PLA₂ induced mice by about 60%, whereas EGCG and ECG lowered the serum CK level by about 40%, indicating the stronger antimyonecrosis activities of dimers than that of the monomers.

3.3. Antihemolysis activities of the four structural elements

As indicated in Figure 3, PT characteristic elements did not show any indirect hemolysis (<5%), whereas PLA₂ alone exhibited strong hemolytic activity, with a hemolysis ratio of 89.05 \pm 3.90%. Both EGCG dimer and ECG dimer significantly decreased the indirect hemolytic activity of PLA₂ (*p* < 0.05), and 31.89 \pm 2.54% and 29.61 \pm 2.07% of neutralization were observed. In contrast, EGCG and ECG monomer exhibited little effects on inhibiting the hemolytic activity of PLA₂, wherein only 18.59 \pm 4.62% and 19.23 \pm 1.39% of neutralization were observed separately.

Table 1 – Effects of PT characteristic structural elements on the lethal toxicity of PLA₂.

	Number of mice	Number of dead mice	Survival ratio (%)	Time until death occurred (h)
PLA ₂	10	10	0	2–6
PLA ₂ –EGCG	10	8	20	6–24
PLA ₂ –ECG	10	9	10	6–24
PLA ₂ –EGCG dimer	10	4	60	6–24
PLA ₂ –ECG dimer	10	5	50	6–24
EGCG	10	0	0	—
ECG	10	0	0	—
EGCG dimer	10	0	0	—
ECG dimer	10	0	0	—

EGC = epicatechin-3-gallate; EGCG = epigallocatechin-3-gallate; PLA₂ = phospholipase A₂; PT = persimmon tannin.

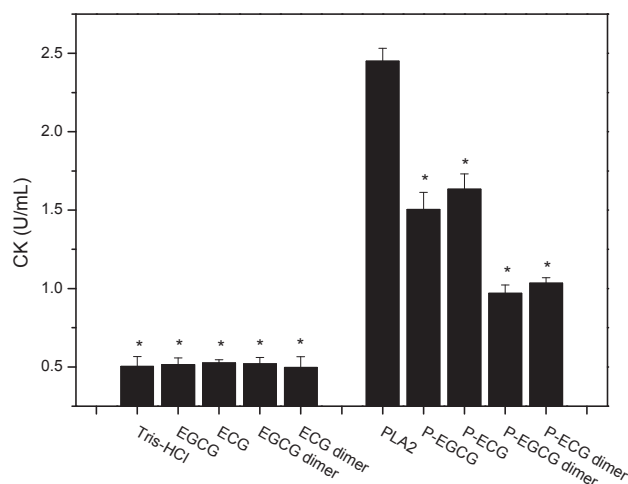


Figure 2 – Effects of persimmon tannin (PT) characteristic structural elements on the myotoxic activity induced by phospholipase A₂ (PLA₂) as measured by creatine kinase (CK) release. Data are expressed as means \pm standard deviation (*n* = 6). **p* < 0.05 as compared to the PLA₂ group.

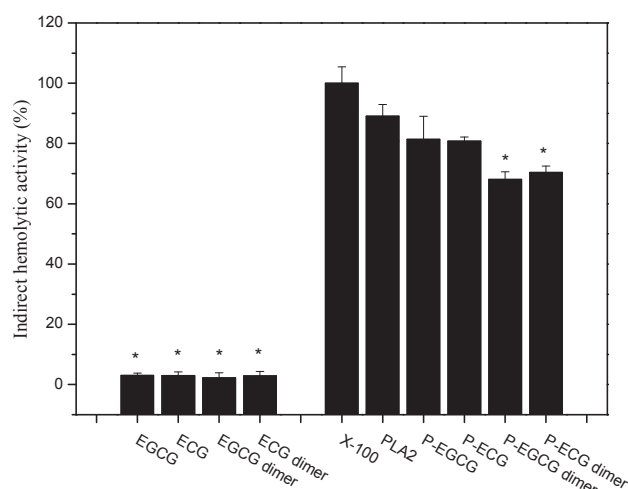


Figure 3 – Effects of PT characteristic structural elements on the indirect hemolytic activity of PLA₂. Data are expressed as means \pm SD ($n = 6$). * $p < 0.05$ as compared to the PLA₂ group. PLA₂ = phospholipase A₂; PT = persimmon tannin; SD = standard deviation.

3.4. Affinities of the four structural elements to PLA₂

For SPR study, the surface of chip was stabilized with PT characteristic elements, and then various concentrations of PLA₂ were injected to the chip. As shown in Figure 4, signal response was observed after injecting the PLA₂ solution, indicating that the interaction occurred. And the interaction was enhanced with the increase of the PLA₂ concentration. Compared with EGCG and ECG monomers, EGCG and ECG dimer generated three times higher signal response with PLA₂, indicating that the interaction between PLA₂ and the dimers was stronger than that with the monomers.

Both association constant (K_a) and dissociation constant (K_d) could be obtained from the SPR curve by fitting the curve with Langmuir equation, and equilibrium dissociation constant (K_D) was calculated from K_a and K_d . As shown in Table 2, the K_D values of EGCG dimer and ECG dimer were 1.69×10^{-7} M and 1.78×10^{-7} M, respectively, whereas those of EGCG and ECG were 2.55×10^{-7} M and 3.34×10^{-7} M, separately, manifesting that the affinity of PT dimers to PLA₂ was higher than that of monomers.

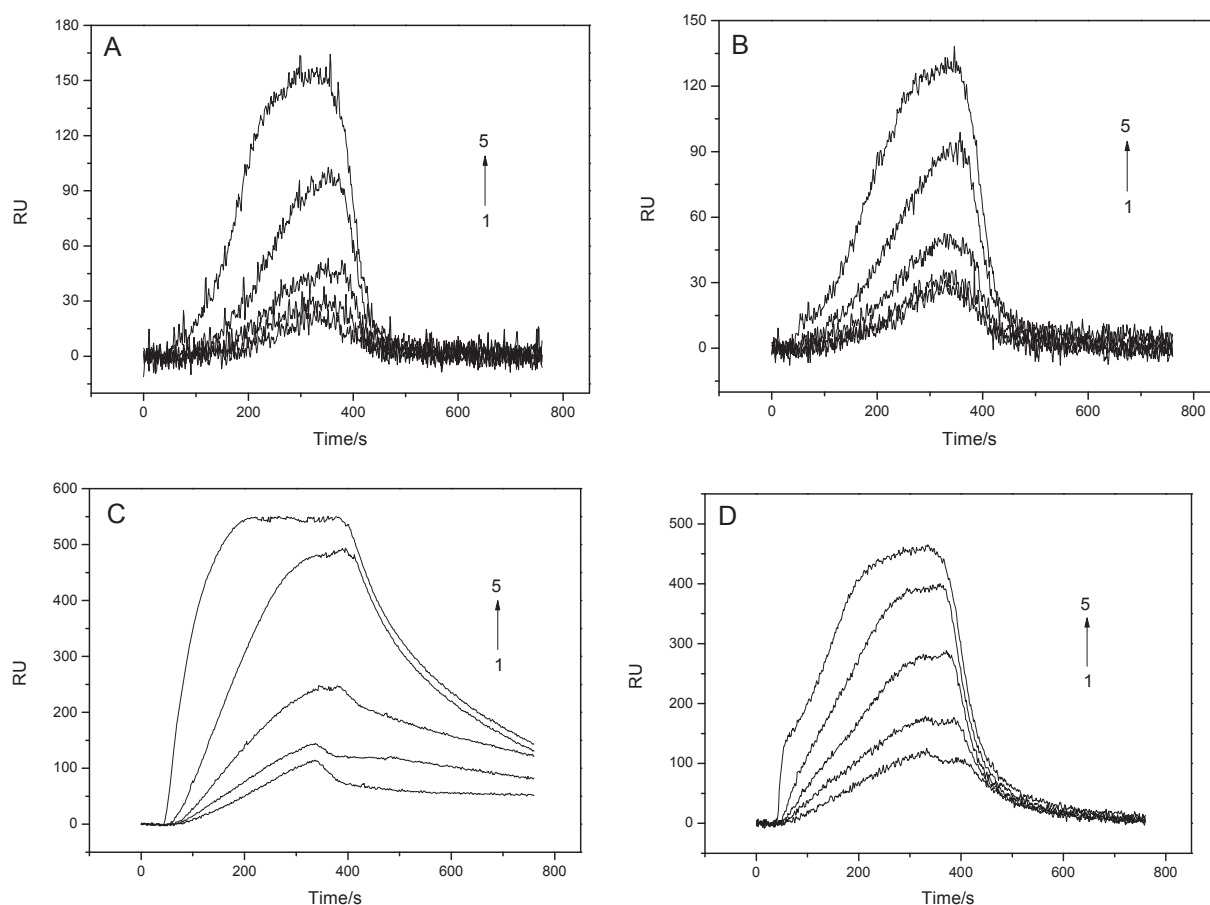


Figure 4 – The binding curves of PT characteristic structural elements with different concentration gradient of PLA₂. 1 \rightarrow 5: 125nM, 250nM, 500nM, 1000nM, 2000nM. (A) EGC G. (B) ECG. (C) EGCG dimer. (D) ECG dimer. ECG = epicatechin-3-gallate; EGCG = epigallocatechin-3-gallate; PT = persimmon tannin.

Table 2 – Dynamic parameters of PT characteristic structural elements binding to PLA₂.

	K _a (1/Ms)	K _d (1/s)	K _D (M)
EGCG	4.99×10^2	1.27×10^{-4}	2.55×10^{-7}
ECG	1.82×10^3	6.07×10^{-4}	3.34×10^{-7}
EGCG dimer	9.81×10^3	1.66×10^{-3}	1.69×10^{-7}
ECG dimer	7.29×10^3	1.30×10^{-3}	1.78×10^{-7}

ECG = epicatechin-3-gallate; EGCG = epigallocatechin-3-gallate; PLA₂ = phospholipase A₂; PT = persimmon tannin.

3.5. Diameters of PLA₂-PT structural elements

The mean diameter of PLA₂ without polyphenols was 4.98 ± 0.18 nm. In the presence of PT characteristic elements, the mean size of PLA₂ continually increased with an increasing concentration of polyphenols (Table 3). As shown in Table 3, the PLA₂-dimer particles formed at high PLA₂/phenol ratios (above 1:20) were significantly larger than that of PLA₂-monomer particles. And it was noteworthy that EGCG dimer induced PLA₂ precipitated at the molar ratio of 1:40. Additionally, larger particle sizes were observed for EGCG dimer than for ECG dimer with PLA₂ at the same molar ratio.

3.6. Effects of four structural elements on the secondary structure of PLA₂

Among the numerous amide bands of the protein exhibited by FT-IR, the amide I band in the range of $1600\text{--}1700\text{ cm}^{-1}$ (representing C=O stretching/hydrogen bonding coupled with COO[−]) and the amide II band (representing NH bending coupled with C–N stretching) have been widely used in studying the secondary structure of protein [11]. As shown in Figure 5, native PLA₂ showed amide I peak at 1654 cm^{-1} and amide II peak at 1543 cm^{-1} . Upon binding with PT characteristic units, the amide I peak showed a significant blue shift from 1654 cm^{-1} to 1630 cm^{-1} , and 9 cm^{-1} of red shift (from 1543 cm^{-1} to 1552 cm^{-1}) was observed in the amide II band, indicating that PT structural units affected the secondary structures of PLA₂ significantly.

To compare the detailed secondary structure changes in PLA₂ after interacting with PT structural elements, amide I of the protein was analyzed using PeakFit software (Figure 6). Based on the curve-fitting of amide I, the secondary structural contents of PLA₂ were obtained. As indicated in Table 4, binding with PT structural units led to a decrease in α -helix and β -turn contents, accompanying an increase in β -sheet and

random coil contents. But addition of different PT structural elements resulted in different degree changes in the secondary structure of PLA₂. EGCG and ECG dimers caused a decrease in the content of α -helix from 42.26% to about 14%, but an increase in β -sheet content from 14.27% to about 63%, whereas adding monomers to PLA₂ led to a decrease in α -helical content to about 18%, and an increase in β -sheet content to about 57%, manifesting that PT dimers affected the secondary structure of PLA₂ more substantially.

4. Discussion

Plant extracts have been traditionally used in the treatment of snakebite envenomations in many countries, especially in rural areas where antivenin is not readily available. Phenolic compounds were one of the main classes of plant extracts that could inhibit snake venoms [12]. Our previous study [7] showed that the PT characteristic elements could inhibit the catalytic activity of PLA₂ potentially, and to explore whether the main structural elements and the characteristic structural units of PT could also inhibit the toxicity of PLA₂, the animal experiments were conducted. Our current data suggested that all four structural elements showed detoxifying effects on PLA₂, but A-type EGCG and ECG dimer exerted significantly more potent antilethality, antimyonecrosis, and anti-hemolysis activities than EGCG and ECG. It was confirmed that the characteristic elements of A-type EGCG and ECG dimer units might also play a vital role in neutralizing the myotoxicity, hemolysis, and lethality of snake venom PLA₂, which extended our previous study. Because both A and B type linkages are existed in PT, it is meaningful to compare the biological activities as well as structural disturbing effects of A type and B type EGCG and ECG dimers on PLA₂. Unfortunately, we were unable to obtain the B type EGCG and ECG dimers from PT, despite our numerous attempts. The preparation of B type dimers of EGCG and ECG is a challenging task, and it requires further work.

It was suggested that phenolic compounds could form complexes with zymoprotein, thus exhibiting the enzyme inhibitory activity [13]. To investigate whether the antitoxic effect of PT structural elements on PLA₂ was correlated with their binding with PLA₂, we first compared the binding affinities of PT characteristic units to PLA₂ using the SPR method. The results showed that PT dimers showed higher affinities to PLA₂ than monomers. However, the SPR method only permits the determination of the interaction between the PT structural elements with PLA₂ in a very short contacting period.

Table 3 – Particle size of PLA₂ prior to and after interacting with PT characteristic structural elements.

Molar ratio (PLA ₂ /phenol)	PLA ₂ -EGCG	PLA ₂ -ECG	PLA ₂ -EGCG dimer	PLA ₂ -ECG dimer
1:0	4.98 ± 0.18	4.98 ± 0.18	4.98 ± 0.18	4.98 ± 0.18
1:1	5.94 ± 0.09	4.73 ± 0.45	6.36 ± 0.32	6.10 ± 0.35
1:10	8.42 ± 0.14	5.94 ± 0.13	8.02 ± 0.11	7.67 ± 0.72
1:20	9.90 ± 0.44	6.88 ± 0.17	1276.48 ± 37.10	1017.87 ± 45.36
1:30	36.50 ± 2.27	13.45 ± 0.74	2303.33 ± 67.04	2132.67 ± 277.80
1:40	525.33 ± 44.34	19.01 ± 4.17	—	2257.33 ± 142.38

ECG = epicatechin-3-gallate; EGCG = epigallocatechin-3-gallate; PLA₂ = phospholipase A₂; PT = persimmon tannin.

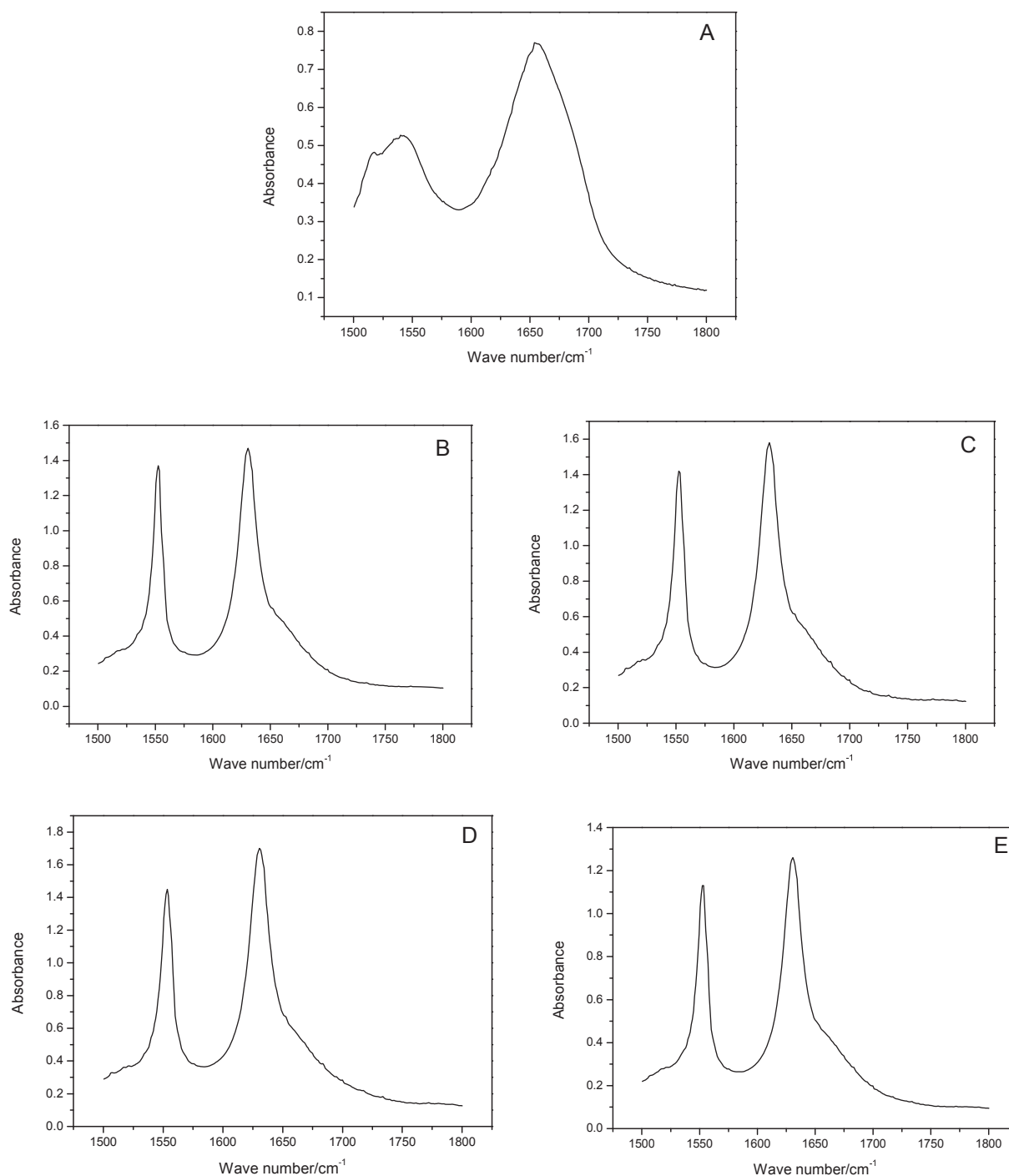


Figure 5 – FT-IR spectra ($1500\text{--}1800\text{ cm}^{-1}$) of PLA_2 prior to and after interacting with PT characteristic structural elements. (A) PLA_2 . (B) PLA_2 –EGCG. (C) PLA_2 –ECG. (D) PLA_2 –EGCG dimer. (E) PLA_2 –ECG dimer. ECG = epicatechin-3-gallate; EGCG = epigallocatechin-3-gallate; FT-IR = Fourier transform infrared spectroscopy; PLA_2 = phospholipase A_2 ; PT = persimmon tannin.

Considering that the interaction of polyphenol on PLA_2 *in vivo* is a complex process, which may last several to dozens of hours, we further prolonged the incubation time of PT structural elements and PLA_2 to 24 hours, and detected the particle size changes of PLA_2 after interacting with PT structural units by DLS methods. The increasing diameter of PLA_2 after

interacting with PT structural units indicated that complexes were formed between them [14]. The difference in the complex size referred to a difference in the binding affinity between protein and polyphenols [15]. PT dimers showed higher affinities to PLA_2 than monomers as expressed by larger particle sizes, which was in line with SPR results. The higher

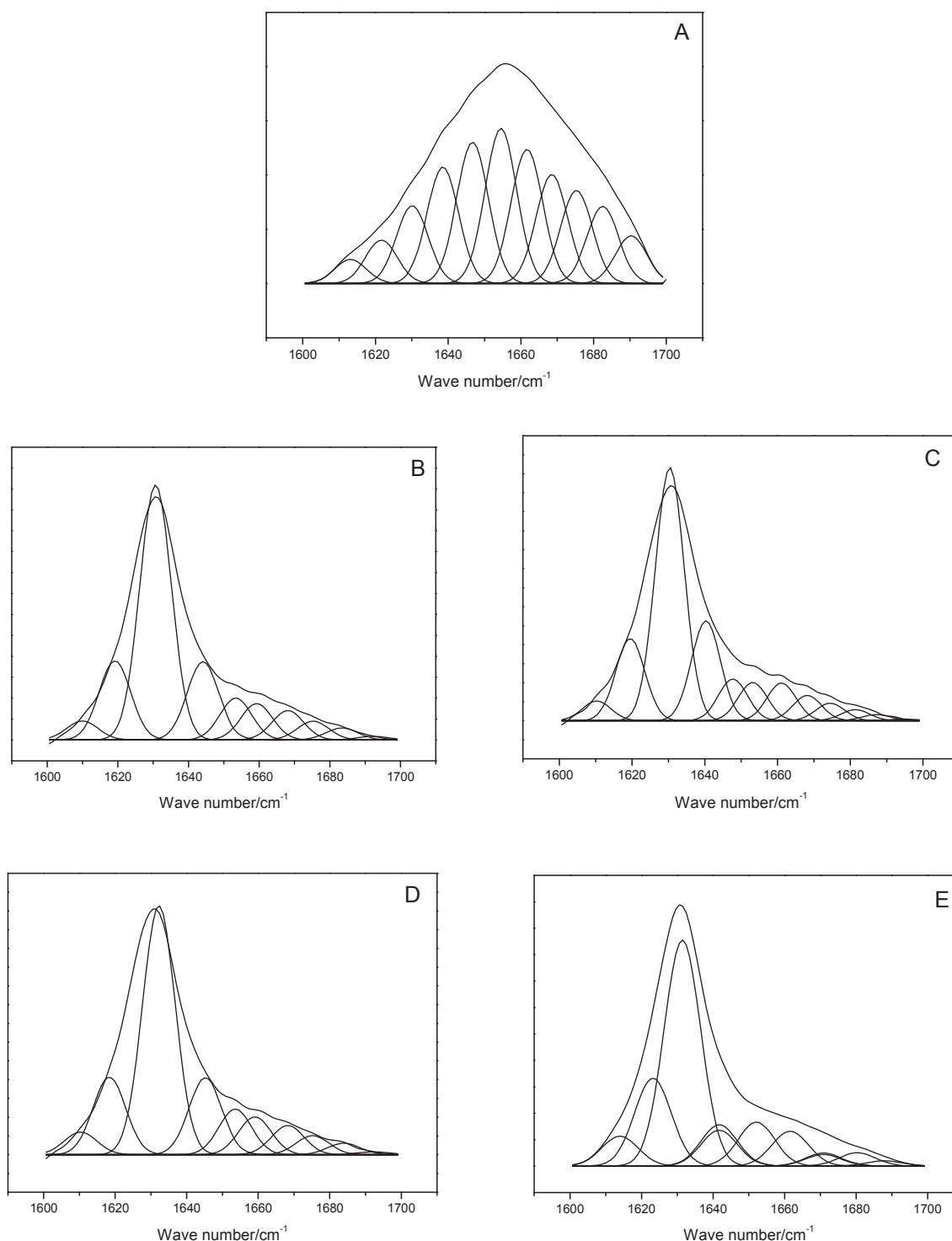


Figure 6 – IR fitting spectra of amide I band of PLA₂ prior to and after interacting with PT characteristic structural elements. (A) PLA₂. (B) PLA₂–EGCG. (C) PLA₂–EGC. (D) PLA₂–EGCG dimer. (E) PLA₂–EGC dimer. ECG = epicatechin-3-gallate; EGCG = epigallocatechin-3-gallate; IR = infrared; PLA₂ = phospholipase A₂; PT = persimmon tannin.

affinities of EGCG and ECG dimer to PLA₂ than that of ECG and EGCG monomers could be explained by the higher molecular weight, the more orthophenolic groups and gallate moieties, as well as the more elongated structure of the former compared with the latter [16]. When paralleling the order of

the affinity of the four PT structural elements to PLA₂ with their detoxifying effects on PLA₂, we found a very similar tendency, indicating that the antitoxic effect of PT structural elements on PLA₂ might correlate with their binding with PLA₂.

Table 4 – Effects of PT characteristic elements on the secondary structures of PLA₂.

	Content of secondary structure of PLA ₂ (%)			
	β-sheet	Random coil	α-helix	β-turn
PLA ₂	14.27	11.43	42.26	32.10
PLA ₂ –EGCG	58.30	14.12	16.12	11.46
PLA ₂ –ECG	56.55	14.26	19.14	10.04
PLA ₂ –EGCG dimer	63.46	14.31	14.19	8.03
PLA ₂ –ECG dimer	63.31	14.18	14.56	7.96

ECG = epicatechin-3-gallate; EGCG = epigallocatechin-3-gallate; PLA₂ = phospholipase A₂; PT = persimmon tannin.

The toxicity of PLA₂ was highly correlated with its spatial structure. In order to illustrate whether the detoxifying effects of PT structural elements correlated with their effects on the structural changes of PLA₂, the secondary structural changes of PLA₂ prior to and after interacting with PT structural units for 24 hours were determined.

Previously, we observed that after incubating with PT elements for 30 minutes, the α-helix content of PLA₂ increased significantly and the β-sheet content of PLA₂ decreased notably [7]. However, a significant decrease in the α-helix content and a notable increase in the β-sheet content of PLA₂ were observed by FT-IR analysis in the current study after extending the incubation time from 30 minutes to 24 hours. It is well known that the interaction between protein and polyphenol is a very complex process and can be affected by numerous factors such as the concentrations of both protein and polyphenol, the ionic strength and pH value of the solution, and the incubation time and temperatures. A slight change in the influencing factor might affect the nature of the interaction between protein and polyphenol [17]. The inconsistent results can be explained by the following reasons. First, because polyphenols were prone to oxidation under neutral or basic conditions in the presence of oxygen [18], prolonging the incubation time from 30 minutes to 24 hours in the current study led to the nonenzymatic oxidation of polyphenols, thus resulting in different secondary structural disturbing effects on PLA₂. Second, during the long period of incubation, molecules of polyphenol or PLA₂ or the polyphenol–PLA₂ complex would undergo rearrangement and self-aggregation, thus exerting structural disturbing effects on PLA₂ that are distinct from those of short-time incubation. Our results suggest that the interaction modes of polyphenols–proteins were not only structure and concentration dependent, but also incubation time dependent. Because the detoxifying process of polyphenol on PLA₂ *in vivo* is a complex process, when elucidating the detoxifying mechanism of polyphenols in the view of the interaction between polyphenols and snake venom, both the existing state of polyphenols under physiological conditions and the interaction time should be considered.

Comprehensively, the four PT structural elements induced conformational and secondary structural changes in PLA₂, of which A-type EGCG and ECG dimer exerted more potent effects on PLA₂ compared with EGCG and ECG. The results were in line with their detoxifying effects on PLA₂, indicating that the spatial structure change of PLA₂ induced by PT elements was probably one of the detoxifying mechanisms of PT.

His48 was reported to be highly conserved in PLA₂, and this residue was essential for the phospholipid hydrolysis activity of PLA₂ [19]. Some pharmacological activities, such as myotoxic, cytotoxic, and hemolytic activities, were probably dependent on the integrity of the catalytic activity [20]. So His was also a key residue in the lethal, myotoxic, and hemolytic activities of PLA₂ aside from the enzymatic activity. Although Lys was reported to be irrelevant to the enzymatic activity of *N. atra* PLA₂, the residue played an important role in lethality as well as anticoagulant, hemolytic, and myotoxic activities [21]. Moreover, Trp in *N. atra* PLA₂ was identified to be not specifically associated with the enzymatic sites or the toxic sites, but still had some relationship with both activities [22]. Another active residue in *N. atra* PLA₂ was Tyr (Tyr3 and Tyr63), which showed important effects on the lethal potency of snake venom [23]. From the secondary structure of *N. atra* PLA₂ [24], it was found that His48, Lys6, Lys115, Trp18, Trp19, and Tyr3 were located in the α-helical structures, whereas Lys56, Lys65, and Tyr63 were located in the β-turn structures. Interestingly, our study showed that PT characteristic elements could decrease both the α-helix and β-turn contents of PLA₂, implying that PT units probably affected the environment of the critical residues, such as His, Lys, Trp, and Tyr, of PLA₂, thus reducing the toxicity of PLA₂. A-type EGCG and ECG dimer affected the secondary structure of PLA₂ more severely, accompanied with stronger effects on the environment of key residues, hence exerting more potent detoxifying capacities than EGCG and ECG.

To conclude, PT characteristic elements (EGCG, ECG, EGCG dimer, and ECG dimer) showed significant neutralizing effects on the myotoxicity, hemolysis, and lethality of Chinese cobra PLA₂. And the detoxifying effects of the four structural elements correlated with their structural disturbing effects well. Our results also suggested that the secondary structure changes of proteins taking place when mixing them with polyphenols differed with the incubation time. Therefore, the interaction time should be considered when elucidating the detoxifying mechanism of polyphenols in view of the interaction between polyphenols and snake venom. Our current work extends our previous work and helps us better understand the antivenom mechanism of PT *in vivo*.

Conflicts of interest

The authors declare that there are no conflicts of interest.

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