Quantitation of Fatty Acids in Milk by Gas Chromatography

AN-BANG WU, WEN-HUI JUAN, HUI-WEN CHENG, CHIN-WEN LIN* AND CHAU-YANG CHEN

Graduate Institute of Pharmaceutical Sciences,
Taipei Medical College, Taipei, Tawian 110, R.O.C.
*Department of Animal Science,
National Taiwan University, Taipei, Taiwan 106, R.O.C.

ABSTRACT

This study investigated a reliable method for quantitation of fatty acids (FA) in milk by gas chromatography. Milk samples were treated with CHCl₃/CH₃OH(2:1, v/v) to extract the fats. A transesterification reaction using fat plus AcCl/CH₃OH(1:10, v/v) as a reagent to prepare fatty acid methyl esters (FAME) was carried out at 60°C for 90 min. The recoveries of FAME derived from standard FA were 98.2±3.2% for the high concentration group; 97.5±4.1% for the low concentration group. When FAME derived from each milk sample was injected into a gas chromatograph, a total of 9 corresponding FA were separated and detected. The FA portion was found to constitute more than 85% of total fat. The content of C4-C12 short-chain FA was also found to be slightly different between the various brands of marketing pasteurized milk and milk powder (reconstituted milk). The content of various FA in long-life milk is essentially similar to pasteurized milk; in flavored milk obvious differences were found due to a variety of additives blended. Raw cow and goat milk contain more short-chain FA than human milk.

Key words: Fatty acids, FAME, milk, gas chromatography.

INTRODUCTION

An important component in milk is lipid. When water is removed from milk, the total solid is classified into two parts, namely fat and solids-non-fat. Generally, the fat content in milk has been reported to be more than $3\%^{(1)}$. Fat contributes to the main flavor of milk, thus various brands of milk sold in market is priced proportionally based on the content of fat. The

milk fat is triglycerides (TG)—the esters formed by glycerol and FA. FA in milk fat are mainly even numbered saturated and unsaturated, normal straight carbon chains. The major saturated FA are⁽²⁾: palmitic acid, stearic acid, myristic acid, butyric acid, while the unsaturated are represented by oleic acid.

There are many well-established methods in separation and determination of FA in milk, including: titration⁽³⁾, spectrophotometry⁽⁴⁾, column chromatography⁽⁵⁾, thin layer chroma-

tography⁽⁶⁾, gas chromatography⁽⁷⁾, and high presssure liquid chromatography⁽⁸⁾. However, gas chromatography has proved to be the most important tool with respect to speed and accuracy⁽⁹⁾.

The objective of this study⁽¹⁰⁾ was to quantify FA in milk and to investigate the differences in the content of FA in pasteurized or reconstituted milk by a modified gas chromatographic method. FA were converted to their methyl esters (FAME) by transesterification in acetyl chloride/methanol solution for better manageability. Finally, a survey was conducted by analyzing 9 FA in various dairy products sampling from local food stores. The quantitative analyses of goat and human milk by GC were also investigated.

MATERIALS AND METHODS

I. Materials

(I). Sources of Milk

Raw cow and goat milk was supplied by Dairy Farm of National Taiwan University. Name brands of pasteurized milk were: Wei-Chuan (W), Tai-Ta (T'), Foremost (F), Kun-Chuan (K). Name brands of milk powder were: Anchor (A), Oak (O), Red Cow (R), Klim (C), Nestle(N) and Wei-Chuan (W). Other milk products were: Wei-Chuan(W), Tai-Nong (T), Kun-Chuan (K) and Foremost (F). These were purchased from local food stores during October, 1986.

(II). Reagents

- 1. Chloroform, methanol, 95% ethanol, so-dium chloride, ammonium hydroxide, diethyl ether: Wako Pure Chemical Industries Ltd. (Japan), extra pure reagent grade. Acetyl chloride: E. Merck, reagent grade.
- 2. Silica gel 60(70-230 mesh ASTM) Art No.77344: E. Merck product.
- 3. Authentic samples: Butyric acid, caproic acid, caprylic acid, capric acid, lauric acid, myristic acid, palmitic acid, stearic acid and oleic acid: E. Merck, reagent grade.

- 4. Reference standards: n-Heptanoic acid and methyl n-heptanoate (MH): E. Merck, reagent grade.
- 5. Internal standards: Ethyl p-hyroxybenzoate (EHB) and butyl p-hydroxybenzoate (BHB): E. Merck, reagent grade.

II. Instruments and Operating Conditions

A gas chromatograph of Hitachi 263-30 Model equipped with a flame ionization detector was used to analyze FAME. The separation was performed on a $2m\times3mm$ ID column packed with 3% OV-101, 80/100 mesh Chromsorb W-HP. Each time $10~\mu l$ of a sample was injected into GC at a programmed column temperature from $80^{\circ}C$ to $280^{\circ}C$ using N_2 as a carrier gas at a flow rate of 40~ml/min.

The GC-MS analyses were performed using a Minnigan TSQ46-C equipped with an EI-Autotune. A column of 15m×0.247mm ID, packed with SE-54, was used. The temperature of injector was maintained at 300°C and the programmed column temperature was adjusted from 60°C to 240°C using He as a carrier gas at a flow rate of 0.2 ml/min.

III. Standard Solutions

(I). FA Stock Solution

FA with carbon chains: C4:0, C6:0, C8:0, C 10:0, C12:0, C14:0, C16:0, C18:0 and C18:1 [Material I.(II).3] were taken in a weight of 5. 0, 10, 20, 40, 50, 80, 100, 150 and 200mg, respectively. All FA were placed in a 10-ml volumetric flask and 1.0ml of a reference standard, n-heptanoic acid (30mg/ml in CHCl₃) was added. The stock solution was diluted with CHCl₃ to the meniscus. The concentrations of FA solution were 0.50, 1.0, 2.0, 4.0, 5.0, 8.0, 10, 15 and 20mg/ml, respectively.

(II). Reference Standard Solution

An amount of 300 mg n-heptanoic acid were weighed and placed in a 10-ml volumetric flask, to which CHCl₃ was added to the meniscus. The concentration of the reference standard was 30 mg/ml.

(III). Internal Standard Solution

One gram of ethyl p-hydroxybenzoate or butyl p-hydroxybenzoate was weighed separately and placed in a 10-ml volumetric flask. By adding CHCl₃ to mark, the internal standard solution was obtained with a concentration of 100 mg/ml.

IV. Reconstituted Milk

Twelve grams of milk powder (each brand) were weighed in a 250-ml flask. Distilled water was added until the solution weighed exactly 100 g. The solution was stirred until the powder was becoming homogeneous.

V. Methods

(I). Extraction of Fat

Folch's method⁽¹¹⁾ was applied by using CHCl₃/CH₃OH (2:1, v/v) to extract liberated FA and TG in milk. Human milk was an exception by adding the solvents and the reagent, CHCl₃/CH₃OH and AcCl/CH₃OH (for methylation) simultaneously without employing the step of extraction.

(II). The Preparation of FAME for GC Analysis

Five grams of a milk sample was weighed accurately in a 10-ml volumetric flask. After the reference standard, p-heptanoic acid (1 ml, 30mg /ml) was added, the sample was diluted with CHCl₃/MeOH (2:1, v/v) to the meniscus. The glycerides were extracted by a separatory funnel for 5 min and centrifuged at 3,000 rpm for 15 min. The lower layer of the solution was collected. Two ml of the solution was pipetted to a flask and washed with 2 ml of 0.0145 mM NaCl solution. Subsequently, 1 ml of the solution was pipetted to a sample vial and 1.0 ml of AcCl/ MeOH (1:10, v/v) was added. The vial was stoppered and sealed with a paraffin film and subject to transesterification in a steam bath at 60°C for 90 min. The sample was ready for GC analysis after 10 μ l of ethyl p-hydroxybenzoate or 30 μ l of butyl p-hydroxybenzoate (an internal standard) was added.

VI. Calibration Curves

The prepartion of FA calibration curves for

GC analysis represented into two groups:

(I). High Concentration Group

One ml of FA stock solution with a concentration of 1.0, 2.0, 4.0, 8.0, 10, 15 and 20 mg/ml was pipetted and transferred separately to a sample vial. One ml of acetyl chloride/methanol reagent (1:10, v/v) was added to the vial, stoppered and sealed with a paraffin film. The sample vial was placed in a steam bath (60°C) for 90 min allowing for complete transesterification. Subsequently, 30 μ l internal standard, butyl phydroxybenzoate solution was added. Ten μ l of the sample was injected into GC; the obtained peak height ratio (the amount of FAME relative to internal standard) was plotted versus the amount of FA. An optimal straight line was obtained by a linear regression analysis.

(II). Low Concentration Group

One ml of FA stock solution with concentration of 0.50, 1.0, 2.0, 4.0 and 5.0 mg/ml was pipetted and transferred separately to a sample vial. After the same treatment as in high concentration group, $10 \mu l$ ethyl p-hydroxybenzoate solution were added as an internal standard. The remaining steps were identical to those in Method VI. (I).

VII. Sample Analysis

(I). Recovery of FAME

The percent recovery of the esters was calculated according to the following formula.

 $\times 100\%$

where

MH=Authentic sample of methyl p-hydroxybenzoate and

IS=Internal standard

(II).Content of FA in Milk

The content of FA in milk was calculated as following:

Content of FA (mg)=
$$\frac{C \times M}{R}$$

where

C=Amount of FA (mg) obtained from the

calibration curve

M=Multiple of dilution

R=The % recovery of methyl ester of the reference standard (n-heptanoic acid)

VIII. Determination of Fat and Total Solid

Roose-Gottieb method⁽¹²⁾ was adopted for quantitation of fat in milk. Five grams milk sample was weighed and placed in a Majonnier tube. Two ml of concentrated ammonia water, 10 ml of 95% ethanol and 10 ml of ethyl ether were added in succession. The mixture was agitated for 30 seconds and allowed to stand vertically for 1 hour. The supernatant was decanted to a 100-ml beaker. The process was repeated by adding 10 ml each of ethyl ether and petroleum ether to the tube. The supernatant was also decanted to the beaker heated at 75°C on a heating plate to evaporate solvents until nearly dry. Subsequently, it was placed in an oven at 90-100 °C for complete dessication until a constant weight was obtained.

RESULTS AND DISCUSSION

Preparation and Characterization of FAME

Prior to the analysis by gas chromatography, all FA must be converted to the derivatives of higher volatility and lower polarity.

The best choice was by methylation of FA to their methyl esters which exhibit the advantages of good yields⁽⁷⁾, shorter retention time and lower boiling points⁽¹³⁾. For the sake of safer and easier manipulation, the reagent acetyl chloride in methanol⁽¹⁴⁾ was used to carry out the transesterification (Scheme 1). The proton released from step 1 could be used as a catalyst to liberate FA from glycerides, immediately followed by step 2-methylation to produce FAME. In a search of the optimum reaction conditions with respect to reaction time (Fig.1) and the concentrations of acetyl chloride (Fig.2), it was found that a complete appareant recovery of FAME could be reached after 80 minutes in a water bath at 60°C with acetyl chloride in methanol (8%, v/v). In the methylation of FA, acetyl chloride in methanol (10%, v/v) with 90 minutes reaction time was actually performed for ensuring the complete recovery of the products.

The structures of the obtained FAME were analyzed by GC-MS: The common features of alkyl cleavage producing O with m/z [C-OCH₃]⁺

59; acyl cleavage producing [OCH₃]⁺ with m/z 31; McLafferty rearrangement producing

O with m/z 74 were observed [CH₃-COCH₃]. +

and correctly identified to be methyl esters. The

1.

$$O = CH_3 - C - C1 + CH_3OH \longrightarrow CH_3 - C - OCH_3 + HC1$$

2

FA'—
$$\overset{\circ}{C}$$
— $\overset{\circ}{C}$ —

Scheme 1. The formation of FAME by transesterification

molecular ions, M⁺ with m/z 102, 130, 158, 186, 214, 242, 270, 298, 296, were identified in sequence to be the methyl ester of butyric acid, caproic acid, caprylic acid, capric acid, lauric acid, myristic acid, palmitic acid, stearic acid, oleic acid, respectively.

Standard Calibration Curves

For the purposes of obtaining accurate GC analysis and the best peak separation, methyl nheptanoate as a reference standard, ethyl p-hydroxybenzoate as double internal standards were carefully tested and chosen. After methylation of the standard solution of FA plus n-heptanoic acid, a sample was injected into GC. A typical gas chromatogram of a standard solution of FAME is shown in Fig.3. A standard calibration curve was completed by plotting the peak height ratio of methyl ester of standard FA to methyl ester of reference standard as abscissa versus the

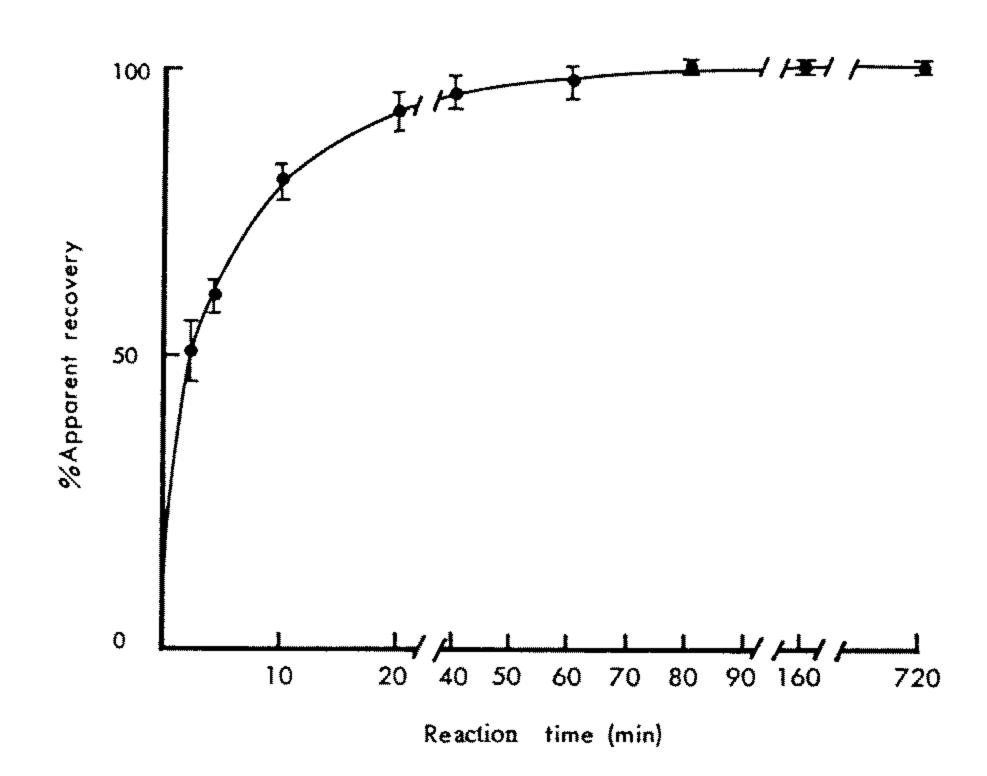
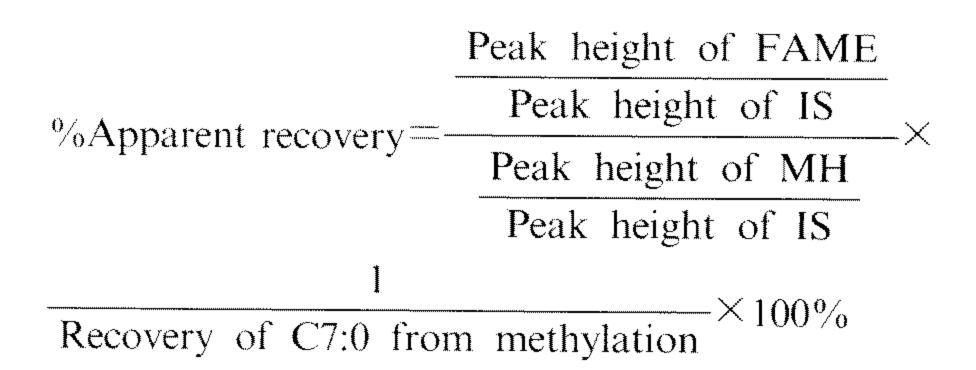


Figure 1. Percent apparent recovery* of FAME as a function of reaction time.



^{*}Expressed as Mean ± S.D. of nine fatty acids

weight (in mg) of FA as ordinate. Nine calibration curves were obtained with regression analyses as shown in Talbe 1.

Recovery of FAME

According to Guy's method⁽¹⁵⁾ of calculating percentage recovery of FA, an internal standard was added to calibrate the loss of the reference standard after methylation. For better accuracy consideration, we divided the FA into

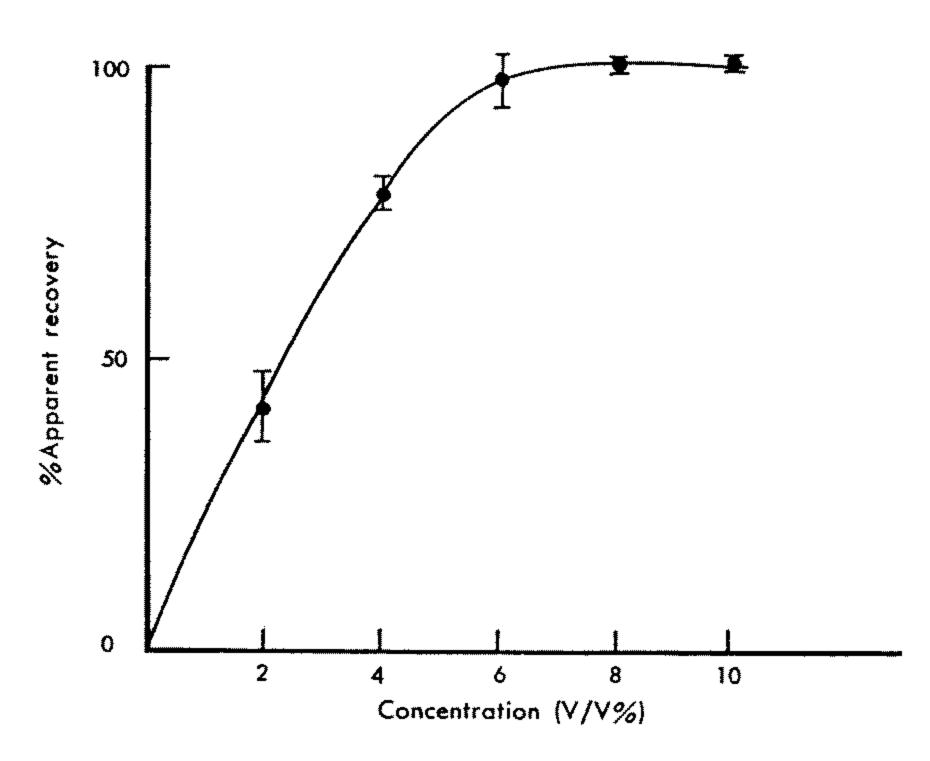


Figure 2. Percent apparent recovery of FAME as a function of acetyl chloride concentration.

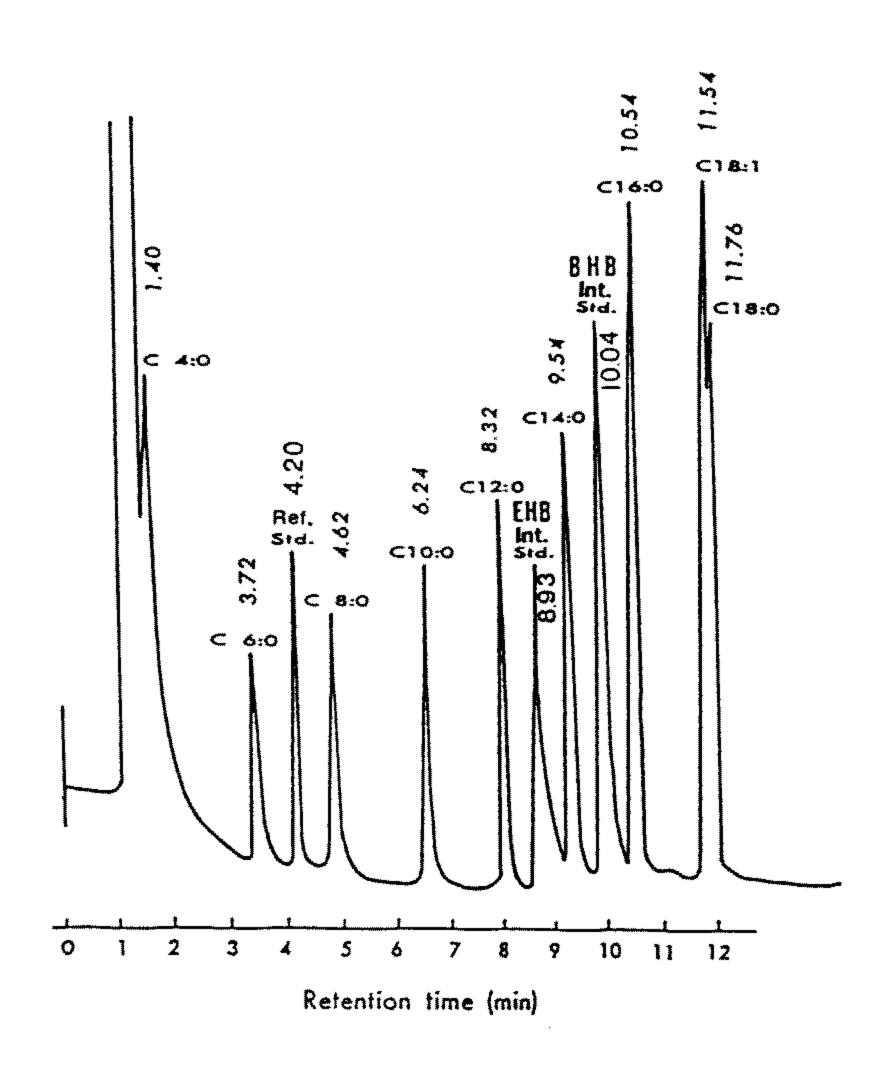


Figure 3. A Gas chromatogram of FAME derived from a standard fatty acids solution.

Table 1. Regression equations obtained from the standard calibration curves of FAME

	Methyl ester of		Correlation	
Calibration curve	fatty acid	Regression equation	coefficient	
			r	
	C14:0	9.673x - 0.078	1.000	
High concentration	C16:0	9.233x - 0.027	0.995	
(1.0-20mg/ml)	C18:0	$9.054x \pm 0.098$	0.997	
	C18:1	9.448x+0.079	0.999	
	C 4:0	0.659x + 0.021	1.000	
	C 6:0	$0.542x \pm 0.010$	0.994	
Low concentration	C 8:0	$0.540x \pm 0.063$	0.998	
(0.50-5.0 mg/ml)	C10:0	$0.644x \pm 0.018$	0.999	
	C12:0	0.694x + 0.014	0.999	

Table 2. The composition of FA in pasteurized milk

Milk sample	\mathbf{W}	T	F	K	Mean \pm S.D.
Total solid ^a	12.0 ± 0.6	12.8±0.7	12.2±0.8	12.0±0.8	12.5 ± 0.7
Total fat ^b	3.4±0.1	3.4 ± 0.2	3.8 ± 0.3	3.6±0.2	3.6±0.2
C 4:0	2.8 ± 0.9	2.8 ± 0.7	3.0 ± 0.2	2.7 ± 0.1	2.8 ± 0.6
C 6:0	1.5 ± 0.2	1.5 ± 0.2	2.0 ± 0.2	1.6 ± 0.2	1.7 ± 0.2
C 8:0	1.5 ± 0.2	1.4 ± 0.2	1.6 ± 0.1	1.6 ± 0.0	1.5 ± 0.2
C10:0	2.2 ± 0.7	2.4 ± 0.4	3.0 ± 0.2	2.4 ± 0.4	2.5 ± 0.5
C12:0	3.0 ± 0.2	3.3 ± 0.4	3.8 ± 0.2	2.9 ± 0.2	3.3 ± 0.3
C14:0	14.3 ± 1.7	19.3 ± 2.6	13.9 ± 0.2	12.8 ± 1.5	15.1 ± 1.7
C16:0	34.6 ± 3.5	34.1 ± 4.1	29.0 ± 0.4	30.4 ± 3.6	32.0 ± 3.2
C18:0	7.6 ± 0.7	7.5 ± 0.2	5.8 ± 0.2	7.2 ± 2.1	7.0 ± 1.1
C18:1	21.4 ± 1.9	22.1 ± 0.9	18.5 ± 0.0	20.2 ± 1.5	20.5±1.3
Total ^c	88.9 ± 10.0	94.4±9.7	80.6 ± 1.7	81.8±9.6	86.4 ± 9.1

a.b. Data expresseed as % w/w sample. (n=5)

high and low concentration groups by adding two internal standards simultaneously-butyl phydroxybenzoate for the calibration of the former; ethyl phydroxybenzoate for the latter. It was found that the recovery of FAME derived from the standard FA was $98.2\pm3.2\%$ (n=5), c. v.=3.3% for the high concentration group; 97. $5\pm4.1\%$ (n=5), c.v.=4.2% for the low concentration group.

Quantitation of FA in Milk

Gas chromatography has been widely used in analyzing FA in dairy and related products up to recently $^{(16-18)}$. By using the present gas chromatographic method to analyze the FA in pasteurized milk of different brands sold at the local food stores, the results are compiled in Table 2. The content of total solid averages $12.5\pm0.7\%$ (w/w); total fat, $3.6\pm0.2\%$ (w/w). A typical gas chromatogram of FAME in milk is shown in Fig.4. Table 3 lists similar analytical

c. Data expresseed as $\frac{9}{6}$ w/w fat. (n=5)

results for various brands of milk powder (12.0g in 88.0 ml H_2O) in the form of reconstituted milk. The content of fat averages $3.4\pm0.6\%$ (w/ w of reconstituted milk). In the comparison of FA content in pasteurized and reconstituted milk, it was found that the former contains

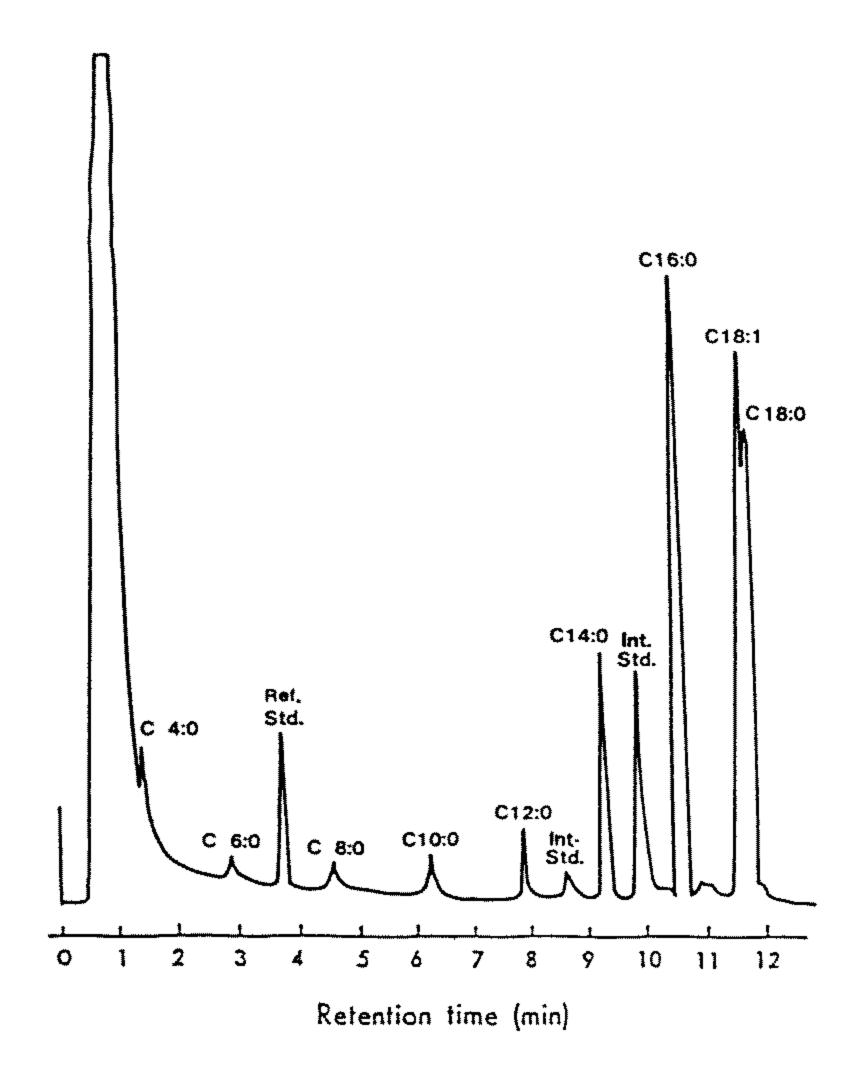


Figure 4. A Gas chromatogram of FAME in milk.

more C6:0, C8:0, C10:0, C12:0, C14:0; and less C4:0 and C18:0 than in the latter. In response to the heightened consumers' sense of demanding better quality of marketing foods today, an exhaustive search for reliable methods in quality control remains necessary. The same demand applies to how to evaluate percentage of blended reconstituted milk in the marketing pasteurized milk. Further examination of Tables 2 and 3 reveals that the variation in contents of shorter C chain (carbon number <10) FA in pasteurized and reconstituted milk seems to provide us a feasible means to evaluate the percentage of reconstituted milk in pasteurized milk sold at market. This area is presently under further investigation.

Furthermore, the GC method was employed to analyze FA in low-fat, long-life and flavored milk sampling from the food stores and the results are given in Table 4; or in human, raw cow and goat milk with results given in Table 5. In general, the percentages (w/w fat) of summing up 9 FA in all brands of the marketing milk or milk powder are similar, about 81 to 90% including 63 to 68% saturated FA and 18 to 22% unsaturated FA. Flavored milk is an

Table 3. The composition of FA in reconstituted milk

Milk sample	W	N	С	О	A	R	Mean ± S.D.
Total solid"	12.0	12.0	12.0	12.0	12.0	12.0	12.0
Total fat ^b	3.4 ± 0.3	3.9±0.4	3.2 ± 0.8	3.3 ± 0.8	3.3 ± 0.9	3.3 ± 0.3	3.4 ±0.6
C 4:0	5.3 ± 0.5	4.9±0.2	5.0 ± 0.2	3.1 ± 0.2	3.8 ± 0.6	5.4 ± 0.2	4.6 ±0.4
C 6:0	1.1 ± 0.2	1.0 ± 0.2	1.2 ± 0.2	1.2 ± 0.2	0.8 ± 0.2	1.2 ± 0.2	1.1 ± 0.2
C 8:0	0.9 ± 0.2	0.6 ± 0.0	0.7 ± 0.0	0.9 ± 0.2	0.6 ± 0.2	0.7 ± 0.0	0.7 ± 0.1
C10:0	2.3 ± 0.5	1.4 ± 0.2	1.7 ± 0.0	2.1 ± 0.8	1.6 ± 0.2	2.5 ± 0.2	1.9 ± 0.4
C12:0	2.5 ± 1.1	2.7 ± 0.2	3.0 ± 0.5	2.8 ± 0.9	2.6 ± 0.4	3.2 ± 1.0	2.8 ± 0.8
C14:0	14.9 ± 0.7	14.3 ± 0.8	15.4 ± 2.0	17.3 ± 1.2	9.5 ± 2.2	16.3 ± 0.2	14.7 ± 1.4
C16:0	34.1 ± 3.2	29.8 ± 3.3	32.5 ± 1.7	37.3 ± 3.5	26.3 ± 1.6	31.7 ± 1.7	32.0 ± 2.7
C18:0	8.2 ± 0.2	7.2 ± 0.4	9.2 ± 0.2	6.4 ± 1.4	6.9 ± 0.2	8.1 ± 0.5	7.7 ± 0.6
C18:1	20.7 ± 0.5	19.3 ± 1.2	24.0 ± 0.5	17.7 ± 3.0	19.6 ± 0.4	21.9 ± 1.5	20.5 ±1.4
TOTAL	90.0 ± 13.7	81.2±6.5	92.7±5.3	88.8±11.4	71.7±6.0	91.0±5.5	86.0 ±8.0

a. Data expresseed as 12.0% w/w (sample equivalent to 1g pasteurized milk)

b. Data expresseed as % w/w sample. (n=5)

c. Data expresseed as $\frac{9}{9}$ w/w fat. (n=5)

Table 4. The composition of FA in various milk products

Milk smple	\mathbf{W}	T	W	K	F	
	(Low-fat)	(Bottled	(Tetra pak	(Flavored)	(Chocolate	(Fruit juice
		long-life)	long—life)		flavored)	flavored)
Total solid ^a	12.1 ± 0.6	11.1 ±0.7	11.4±0.6	13.6±0.5	12.9 ± 0.7	12.7 ± 0.8
Total fat ^b	1.9±0.3	3.2±0.2	3.3 ± 0.4	3.7 ± 0.3	2.7±0.3	2.8±0.6
C 4:0	3.9±0.4	4.6±0.3	4.5±0.9		1.7±0.0	
C 6:0	1.6 ± 0.4	0.7 ± 0.0	0.8 ± 0.0	***************************************	***************************************	0.2 ± 0.1
C 8:0	1.6 ± 0.0	0.4 ± 0.3	0.7 ± 0.0		3.0 ± 0.7	2.3 ± 0.6
C10:0	2.0 ± 0.0	1.8 ± 0.5	0.6 ± 0.0		1.6 ± 0.7	1.4 ± 0.9
C12:0	2.5 ± 0.0	2.8 ± 0.3	2.0 ± 0.2		34.6 ± 2.3	30.3 ± 5.6
C14:0	14.1 ± 3.2	13.2 ± 0.5	13.4 ± 0.2	1.4 ± 0.0	16.9 ± 1.7	15.2 ± 2.7
C16:0	35.1 ± 3.2	31.9 ± 1.0	33.5 ± 0.5	17.7 ± 1.2	8.1 ± 0.7	9.0 ± 2.9
C18:0	5.6 ± 1.6	7.6 ± 0.3	7.5 ± 0.2	26.2 ± 1.4	2.1 ± 0.0	2.2 ± 0.6
C18:1	19.6±2.0	21.6 ± 0.8	21.4 ± 0.7	38.5 ± 2.1	5.9 ± 0.3	6.3 ± 1.8
$TOTAL^c$	86.1±11.3	83.6±4.0	84.4 ± 2.7	83.8±4.7	70.1 ± 6.4	67.0 ± 15.4

a.b. Data expresseed as % w/w milk sample. (n=5)

Table 5. The composition of FA in human milk, raw cow and goat milk

Fatty acid *	Human milk ^a	Cow milk ^b	Goat milk ^h 1.82 ± 0.25	
C 4:0		2.26±0.28		
C 6:0		1.08 ± 0.04	0.44 ± 0.10	
C 8:0		1.04 ± 0.04	0.41 ± 0.06	
C10:0	1.03	1.43 ± 0.13	1.26 ± 0.18	
C12:0	6.93	1.62 ± 0.10	1.81 ± 0.03	
C14:0	3.32	6.50 ± 0.42	0.48 ± 0.04	
C16:0	14.31	14.30 ± 0.88	14.00 ± 0.57	
C18:0	4.76	3.02 ± 0.9	3.26 ± 0.35	
C18:1	10.32	8.60 ± 0.53	9.28 ± 0.23	
Total	40.67	39.85±2.61	32.76 ± 1.81	

^{*} Expressed as mg/g sample

exception due to saturated, unsaturated FA or fruit juice added during processing. The contents of FA in cow or goat milk were also found to be approximately equal, while shorter C chain (C number < 10) FA are not found (trace amount) in human milk. This study extends the potential application of GC to the FA analysis of

commerical milk samples and is a reliable and reproducible method.

REFERENCES

1. Lin, C.W. 1993. Characteristics and Functions of Dairy Products. First Edition. p.36

c. Data expresseed as $\frac{9}{9}$ w/w fat. (n=5)

a. One determination

b. Mean \pm S.D. (n=5)

- Hua-Hsian-Yuan Publisher, Taipei.
- 2. Hilditch, T.P. and Meara, M.L. 1944. Human-milk Fat. Biochem. J. 38:29.
- 3. Howard, K. and Max, E.R. 1974. A Gas Liquid Chromatography Assay for Plasma Free Fatty Acids. J. Chromatogr. 88:253.
- 4. Duncombe, W.G. 1983. The Colorimetric Micro-determination of Long Chain Fatty Acids. Biochem. J. 88:7.
- 5. Eric, N.C., Graeme, F.D. and Owen, A.J. 1983. A Method for the Free Fatty Acids Determination of Individual Free Fatty Acids in Milk by Ion Exchange Resin Adsorption and Gas Liquid Chromatography. J. Dairy Res. 50:321.
- 6. Duthie, A.H. 1965. New Method for Extraction of Milk Phospholipid. J. Dairy Sci. 48: 170.
- 7. Jacqueline, M.L., Trafford, D.J.H. and Makin, H.L.J. 1981. A Rapid and Simple Method for the Esterification of Fatty acid and Steroid Carboxylic Acid Prior to Gas Liquid Chromatography. Clin. Chim. Acta. 111:91.
- 8. William, W.C., Kevin, C. and Raymond, C. N. 1984. Preparative Separation of Milk Fatty Acid Derivatives by High Performance Liquid Chromatography. J. Chromatogr. 298:513.
- 9. Jensen, R.G., Quinn, J.G., Carpenter, D.L. and Ampugna, J.S. 1967. Gas Liquid Chromatographic Analysis of Milk Fatty Acids. J. Dairy Sci. 50:119.
- 10. Juan, W.H. 1987. Determination of Fatty Acids by Gas Chromatography and Spectrophotometric Properties of Whey Protein in Milk. Master Thesis, Taipei Medical College.

- 11. Folch, J.M., Lees, G.H. and Sloane, S. 1957. A Simple Method for the Isolation and Purification of Total Lipid From Animal Tissues. J. Biol. Chem. 226:497.
- 12.A.O.A.C. 1990. Chapter 33. Dairy Products. Milk. In Official Methods of Analysis, 15th Edition, pp.804-835. Assoc. Off. Anal. Chem. Washington. D.C., U.S.A.
- 13. Ulrich, H., Harold, R. and Gunther, G. 1973. Gas Chromatography-Mass Spectrometry of C1-C20 Fatty Acid Benzyl Esters. J. Chromatogr. 87:481.
- 14. Howard, K. and Max, E.R. 1974. A Gas Liquid Chromatographic Assay for Plasma Free Fatty Acids. J. Chromatogr. 88:253.
- 15. Guy L. and Claude, C.R. 1984. Improved Recovery of Fatty Acid through Direct Transesterification without Prior Extraction or Purification. J. Lipid Res. 25:1391.
- 16. Ballesteros, E., Cardenas, S., Gallego, M. and Valcarcel, M. 1994. Determination of Free Fatty Acids in Dairy Products by Direct Coupling of A Continuous Preconcentration lon-Exchange-Derivatization Module to A Gas Chromatograph. Anal. Chem. 66 (5):628.
- 17. Dotson, K.D., Jerrell, J.P., Picciano, M.F. and Perkins, E.G. 1992. High-Performance Liquid Chromatography of Human Milk Triacylglycerols and Gas Chromatography of Component Fatty Acid. Lipids. 27(11):933.
- 18. Matter, L. 1992. Determination of the Animal of Origin of Dairy Products and Raw and Cooked Meats by GC Analysis of the Fatty Acid Methyl Esters (FAME) Obtained by Transesterificaion. J. High Resol. Chromatogr. & Chromatogr. Commun. 15(8):514.

以氣相層析法測定牛奶中脂肪酸含量

吳安邦 阮文惠 鄭慧文 林慶文* 陳朝洋

臺北醫學院 藥學研究所 *國立臺灣大學 畜產學研究所

摘 要

本研究係以氣相析法探討牛奶中脂肪酸定量之可行性。牛奶以氯仿/甲醇(2:1, v/v)萃取脂肪,以氯化乙醯/甲醇(1:10, v/v)在60°C經90分鐘後,轉酯反應完成,可製成脂肪酸甲基酯。以脂肪酸標準品製成甲基酯其回收率為高濃度組,98.2±3.2%;低濃度組97.5±4.1%。將檢樣注入氣相層析儀分析

結果:各類牛奶檢樣中檢出九種主要脂肪酸,佔總脂肪量85%以上。市售主要廠牌之鮮奶與奶粉中, C4-C12短鏈脂肪酸含量略有差異。保久乳中各類脂肪成分大致與鮮奶相同。而調味乳依廠牌不同, 皆摻入添加物致脂肪酸含量有顯著差異。牛奶和羊奶較人奶含有較多短鏈脂肪酸。